

5-15-2019

Vulnerability of juvenile clams to predation by shore crabs

Megan Dethier

Katie Dobkowski

Bates College, kdobkows@bates.edu

Alifaire Noreen

Malise Yun

Abigail Moosmiller

Follow this and additional works at: https://scarab.bates.edu/faculty_publications

Recommended Citation

Dethier, M.N., Dobkowski, K., Noreen, A., Yun, M. and Moosmiller, A., (2019) Vulnerability of juvenile clams to predation by shore crabs. *Aquaculture*. 506, 350-354. <https://doi.org/10.1016/j.aquaculture.2019.03.070>

This Article is brought to you for free and open access by the Departments and Programs at SCARAB. It has been accepted for inclusion in All Faculty Scholarship by an authorized administrator of SCARAB. For more information, please contact batesscarab@bates.edu.

Vulnerability of Juvenile Clams to Predation by Shore Crabs

Megan N. Dethier. Biology Department and Friday Harbor Laboratories, University of

Washington, Seattle, WA

Katie Dobkowski. Biology Department and Friday Harbor Laboratories, University of

Washington. Current address: Biology Department, Bates College, Lewiston, ME

Alifaire Noreen. Friday Harbor Laboratories. Current address: Western Washington University,

Bellingham, WA

Malise Yun. Biology and Anthropology Departments, University of Washington, Seattle, WA

Abigail Moosmiller. School of Aquatic and Fishery Sciences, University of Washington, Seattle,

WA

Running title: Juvenile clam vulnerability to crab predation

Keywords: juvenile clams; predation; crabs; size refuge; shell crushing

18 **Abstract**

19 Predation, especially by crabs, is a common source of mortality for natural and farmed
20 populations of intertidal clams. Consumption of juvenile “seed clams” forces aquaculturists to
21 try to exclude predators and/or raise juveniles in hatcheries until they can reach a size refuge.
22 We ran a variety of lab experiments assessing vulnerabilities of juvenile clams to small,
23 common shore crabs (*Hemigrapsus* spp.). Crabs <1 cm width can consume hardshell Manila
24 clams larger than those normally used for “seed”, and can readily crush even larger softshell
25 *Mya* clams. We suggest that using netting to prevent consumption by shore crabs is not
26 practical given that smaller individuals can fit through mesh openings. Raising seed in
27 hatcheries until they are 10-15 mm will provide a size refuge from shore crabs, but not larger
28 cancrid crabs. Farming on beaches with little habitat (e.g., cobbles) for shore crabs can likely
29 reduce juvenile clam mortality. A better understanding of predation threats to commercially
30 important clams is critical, especially as the invasion of the green crab *Carcinus* to Washington
31 shorelines further threatens survival of juvenile clams.

32

1. Introduction

Aquaculture is an increasingly important contributor to food production and the global economy, with the worldwide farming of shellfish increasing by 650% in the past 30 years (Saurel, Ferreira, Cheney, Suhrbier, Dewey, Davis, Cordell, 2014). While some aspects of shellfish farms raise ecological and aesthetic concerns (Bendell, 2015; Munroe, Kraeuter, Beal, Chew, Luckenbach, Peterson, 2015), culturing bivalves (oysters, mussels, and clams) can provide ecosystem services as well as a valuable protein source. These services include reducing water turbidity and excess nitrogen (Higgins, Stephenson, Brown, 2010), sequestering carbon in shells, and even reducing wave energy that erodes shorelines (Peterson, Grabowski, Powers, 2003).

A substantial challenge to the success of both farmed and natural populations of shellfish is reducing loss to predators. Diverse consumers such as seastars, carnivorous snails (whelks and moonsnails), crabs, birds (shorebirds and diving ducks), and fishes (bony fishes and rays) can all reduce densities of adult clams (Beal, 2006; Bendell, 2015; Beukema, Dekker, Philippart, 2010; Munroe, McKinley, 2007; Myers, Baum, Shepherd, Powers, Peterson, 2007). Recently settled (<1 mm) clams of various species are also vulnerable to mortality from a range of predators including crangonid shrimp, juvenile crabs including hermit crabs, grapsids, and portunids, nereid polychaetes, and perhaps even nematodes (Beukema, Dekker, 2005; 2014; Hunt, Scheibling, 1997; Olafsson, 1989; Walton, MacKinnon, Rodriguez, Proctor, Ruiz, 2002; Whitton, Jenkins, Richardson, Hiddink, 2012; Williams, 1980). Slightly larger clams (2-5 mm) can be eaten by other predators including nemertean worms, numerous crab species, and

mummichog and other fishes (Beal, Vencile, 2001; Hunt, Mullineaux, 2002; Peterson, Summerson, Huber, 1995; Walton, MacKinnon, Rodriguez, Proctor, Ruiz, 2002).

Large losses of clams at shellfish farms to predators, especially crabs, leads to expensive mitigation efforts to increase clam survival (Peterson, Summerson, Huber, 1995). “Seed clams” (usually 1-15 mm) have higher survival following outplanting when they are protected from predators by plastic nets (reviewed by Munroe, Kraeuter, Beal, Chew, Luckenbach, Peterson, 2015) or by coarse substrate such as gravel added to the sediment surface (Glaspie, Seitz, Ogburn, Dungan, Hines, 2018; Peterson, Summerson, Huber, 1995; Ruesink, Freshley, Herrold, Trimble, Patten, 2014). Coarser substrates may give clams surfaces to attach to for stability (Becker, Barringer, Marelli, 2008) or make it harder for predators to dig (Arnold, 1984; Liu, Wang, Lu, Hu, Su, Liu, Zhu, 2019). Ironically, in some cases adding coarser substrate may encourage residence of small crabs, which can consume seed clams (Smith, Langdon, 1998). However, netting and sediment amendment will not increase survival if a key mortality source is small predators that are undeterred by the large mesh of most anti-predator netting.

The Manila clam, *Ruditapes philippinarum* (Adams & Reeve, 1850) (also known as *Venerupis philippinarum*, *Venerupis japonica* and *Tapes japonica*), is an important fisheries species in the Caribbean, Mediterranean, North Atlantic Ocean and North Pacific Ocean. In 2014, global production was 4,000,000 tons (Cultured Aquatic Species Information Programme 2005: http://www.fao.org/fishery/culturedspecies/Ruditapes_philippinarum/en). Manilas were introduced to the US west coast in the late 1930s and are now an important farmed species in Washington State (Smith, Langdon, 1998). Another introduced species that is less broadly farmed in Washington is the softshell clam *Mya arenaria* Linnaeus, 1758, which is an

economically critical shellfish species in the northeast US (Glaspie, Seitz, Ogburn, Dungan, Hines, 2018). For Manilas, as many as 100,000 larvae per m² may settle into natural sediment (Tezuka, Kamimura, Hamaguchi, Saito, Iwano, Egashira, Fukuda, Tawaratsumida, Nagamoto, Nakagawa, 2012) but <1% survive to harvestable size (40 mm in length; e.g., Williams, 1980). This very high post-settlement mortality could stem from abiotic factors such as salinity or temperature variation, or biotic factors such as predation and competition for resources (Dethier, Ruesink, Berry, Sprenger, 2012; Hunt, Scheibling, 1997; Williams, 1980).

As part of a larger study on sources of mortality to juvenile clams on Washington shorelines, we investigated the vulnerability of juvenile clams to crab predation, and how this is affected by clam size and species. Here we focus on the potential role of shore crabs, *Hemigrapsus nudus* (Dana, 1851) and *H. oregonensis* (Dana, 1851). These small (< 4 cm width) crabs are abundant on shorelines from Alaska to the Gulf of California, inhabiting numerous habitat types (mud to bedrock) but especially soft-sediment shorelines where cobbles or debris give them protection at low tide (Harger, 1972; Kozloff, 1993; Low, 1970). The two species are largely sympatric although *H. oregonensis* tends to prefer somewhat more wave-protected shores (Harger, 1972). Unlike other decapods such as cancrid crabs or invasive green crabs (*Carcinus maenas*) that are known consumers of shellfish, Quayle (1988) dismissed *Hemigrapsus* as important predators on shellfish; he noted that these species are commonly found under clusters of oysters but that “they are scavengers and do no harm to oysters” (p. 93). Other authors have considered them to be herbivorous or omnivorous (Knudsen, 1964; Yamada, Boulding, 1998).

While *Hemigrapsus* spp. are not usually considered to be significant intertidal predators, we hypothesized that they could contribute to high field mortality of newly settled clam spat and

juveniles. We were particularly interested in when clams reach size refuges from these predators. If small crabs that are not excluded by shellfish farm netting can consume seed clams, then seed clam sizes may need to be increased to reduce field mortality.

2. Methods

2.1 Field data on predator densities

Work reported elsewhere (Dethier, Kobelt, Yiu, Wentzel, Ruesink, 2019) at study beaches around Washington State quantified abundances of large (> 5 cm) cancrid crabs, but mesh size of the Fukui traps used was too large (12 mm) to retain smaller predators such as shore crabs. We counted shore crabs in quadrats along 50 m horizontal transects in the mid-shore (+1 m above Mean Lower Low Water (MLLW)), but found that this method also did not effectively quantify these mobile predators; at low tide they tend to cluster under cobbles or debris (e.g. logs, netting) that were not necessarily found along the transect. We thus present data from other studies showing the ranges of abundances of shore crabs on Washington beaches. These data (Dethier, Schoch, 2005) come from monitoring surveys of over 30 sites and up to 15 years. In each case, site-date values are counts of crabs averaged over ten 0.25 m² quadrats along horizontal transects at either MLLW or +1 m MLLW.

2.2 Lab experiments with crabs consuming clams

Experiments were conducted in indoor sea tables with running seawater (at 10.5-12.3°C) at the Friday Harbor Laboratories (FHL), Washington (48° 32' 45" N 123° 00' 45" W) during 2017. We worked with juvenile clams of two commercially important species: *Ruditapes philippinarum* (Manila clams) that were obtained from Taylor Shellfish at 1 or 2 mm shell length

or field-collected at larger size; and softshell clams *Mya arenaria* that were collected at various field sites. All clams were kept in sea tables while being fed Shellfish Diet 1800 (Instant Algae®). Shore crabs *Hemigrapsus nudus* and *H. oregonensis* were collected from under intertidal cobbles on the FHL beach. For all individuals used in experiments we measured both carapace width and dactyl length (inner surface) of the larger claw. Yamada and Boulding (1998) show that female crabs have smaller claws; we did not track sex as we assumed that the key parameter in ability to consume clams was claw size regardless of sex. Crabs were fed ulvoid algae if kept in lab for more than several days, but were always starved for 24 hours before experiments. Each individual crab was only used in one experiment.

We tested the abilities and preferences of *Hemigrapsus* spp. of different sizes to consume clams of different sizes. Feeding trials were run by placing individual crabs in small (ca. 10x10x10 cm) lidded plastic containers with mesh sides to allow water flow. Pilot studies quantifying predation rates with or without sediment in the containers showed no consistent differences, so experiments were run without sediment to improve the consistency of data collected. To determine how large a clam could be consumed by crabs, a single measured (length) clam was placed in a container with a crab. After 24 hours, the clam was recorded as intact or crushed and consumed, and the crab was measured and released. Most experiments were run with small Manila clams, but some parallel trials were run with juvenile *Mya arenaria*.

To determine relative predation rates, individual crabs were given 10-20 small measured Manila clams, either all one size (to quantify consumption rates) or of a range of sizes (to quantify preferences). Numbers and sizes consumed were quantified after 24 hours.

3. Results

3.1 Predator densities at field sites

In quadrats at eight sand-pebble beach sites (Dethier, Kobelt, Yiu, Wentzel, Ruesink, 2019), *Hemigrapsus* spp. were sparse (overall average of less than one per 0.1 m² quadrat) and tended to be very small (average size of 5 mm) individuals that could hide effectively along the transect in clam holes or under pebbles. More and larger (up to 3 cm) individuals were seen when cobbles or debris were overturned at almost any intertidal elevation. Other beaches in the state (see Methods) had densities of *H. oregonensis* ranging from 4-40/m² in the mid shore (at ~+1 m, N = 82 site-date combinations) and 0.4 to 20 in the low shore (MLLW, N = 186 site-dates). Highest densities were found where cobbles provided shelter on sand or gravel beaches. *H. nudus* is more common higher on the shore than at either of these transect elevations (Dethier, pers. obs.) and tends to be more abundant at higher-energy sites (Harger, 1972; Kozloff, 1993). Yamada and Boulding (1996) reported densities of *Hemigrapsus* spp. up to 300/m² on optimal gravel-cobble beaches. Low (1970) found that densities of both species were very low (ca. 0.1/m²) on bedrock shores and on plain sand and mud beaches with little 3D structure, but as high as 500/m² on beaches with loose rocks on the surface. The amount of cover, especially from cobbles, was the best predictor of abundance of both crab species. Diverse vertebrates prey on them, including raccoons, gulls, a variety of fishes, and many diving ducks, so significant populations are only found where they have refuges during both low and high tides (Low 1970).

3.2 Lab experiments with crabs

Our feeding observations showed minimal differences in predation rates or preferences between *Hemigrapsus nudus* and *H. oregonensis*, so we pooled results from these two taxa. Both crab species ranged in sizes of carapace width from approx. 5 to 25 mm and dactyl length from 0.5 to 7.7 mm. The tight correlation ($r^2 = 0.88$, $N = 66$) between carapace and dactyl sizes suggests that at least in our sampled mixed-gender and mixed-species population, there was not a substantial difference among sexes or species in relative claw sizes (Suppl. Fig. 1). Here we report results by dactyl length, assumed as the more relevant parameter for ability to break a clam shell.

Both shore crab species showed high individual variance in behavior in terms of how many small (2 mm) clams they ate in a given 24 hour period. Crabs of every size tested ate from 0 to 10 (all) clams per day, with no discernable pattern to these differences ($N = 126$ trials; Suppl. Fig. 2). We thus focus our analyses on how big a clam *could* be eaten by a particular size crab, and on relative rates of predation as crab and clam sizes were varied. We found that even the smallest *Hemigrapsus* tested (~1 cm carapace width, with ~1 mm dactyls) could crush 2 mm Manila clams. When offered the thinner-shelled *Mya* clams, small crabs could crush a much larger individual compared to Manila clams (Figure 1); a 12 mm (carapace width) crab with a 2.5 mm dactyl could crush a *Mya* almost its size (11 mm shell length). Larger Manila individuals (ca. 10 mm) could only be crushed by the larger (~25 mm carapace) shore crabs.

Because we had few juvenile *Mya* available for experiments, predation rate experiments with shore crabs were run only with Manila clams. Small crabs (dactyl < 4 mm) readily ate small (<5 mm) clams, sometimes consuming all 10 offered within 24 hours (Fig. 2). The data from

medium-sized crabs (dactyl 4-6 mm; carapace 15-20 mm) show a pattern of the crabs eating fewer large clams than medium or small ones in the same period of time. Large crabs readily ate all sizes of clams including the larger individuals (Fig. 2).

When medium to large (mean dactyl length 6.8 mm: carapace width ~20mm) shore crabs were offered a choice of 5 individuals of each of the 3 size classes of clams, they showed no preferential consumption for any size (Suppl. Fig. 3), eating ca. half of all the clams regardless of size.

4. Discussion

Crabs inhabiting the intertidal zone on Washington state shorelines (e.g., *Hemigrapsus* spp.) or foraging there at high tide (cancrids; also majids, *Telmessus*: pers. obs.) likely constitute significant sources of mortality for clams, as has been shown in numerous other systems (see Introduction, also Boulding, 1984). Unfortunately, it is difficult to precisely quantify abundances of such mobile predators. Other investigators have tried to quantify densities via snorkeling at high tide (Grosholz, Ruiz, Dean, Shirley, Maron, Connors, 2000) or benthic trawling (Glaspie, Seitz, Ogburn, Dungan, Hines, 2018). Crab abundances can be very patchy, e.g. due to local hypoxia (Altieri, 2008), or the presence of refuges from their predators, e.g. from gulls (Beal, 2006). The *Hemigrapsus* shore crabs we worked with are broadly present on Washington shorelines except at the most wave-exposed beaches, and can be present in hundreds per m² in areas where cobbles or debris provide low-tide refuges (Low, 1970). Thus, their predatory impact on juvenile clams could be substantial.

We found that in the lab, even very small *Hemigrapsus* spp. can crush and consume not only juvenile softshell clams (*Mya*) but hardshell Manilas. Clams <5 mm were consumed by even the smallest shore crabs tested (carapace width 5-10 mm). Adult shore crabs (carapace >20 mm) readily ate 8 mm clams, even hardshell Manilas. Some crabs tended to eat fewer large clams than smaller ones in the same period of time; this could result either from longer handling times (not measured), or satiation when eating the larger prey. Crabs were very individualistic in terms of feeding rates; under identical conditions (starved, then offered clams for 24 hours in lab), some ate nothing while others consumed all the clams available.

As in other studies, consumption by crabs clearly varies not only with clam size but with shell morphology (see also Boulding 1984), with the thinner-shelled *Mya* more vulnerable to crushing. When clam shells are weakened by acidified porewater conditions, (Glaspie, Longmire, Seitz, 2017; Green, Waldbusser, Reilly, Emerson, O'Donnell, 2009), this vulnerability to shell-crushing predators increases. Some experiments have also shown that crab feeding behavior can be directly affected by pH (Glaspie, Longmire, Seitz, 2017), complicating our ability to predict the effects of this increasingly relevant abiotic stressor.

Our data thus contribute to the literature on the ability of clams to reach size refuges from their predators. In our system, Manila clams, despite their hard shells, do not reach a size refuge from the very common intertidal shore crabs until they are >12 mm. Although we did not run experiments with adult clams, it is likely that there is no size refuge for Manilas from the cancrid crabs that were present at most of our sites (Boulding, 1984; Boulding, LaBarbera, 1986; Dethier, Kobelt, Yiu, Wentzel, Ruesink, 2019; Yamada, Boulding, 1996; 1998). Small (20 mm) *Cancer oregonensis*, for example, can consume oysters over 30 mm length (Yamada,

Metcalf, Baldwin, 1993). Survival of farmed Manila clams to harvestable size thus must rely on netting to exclude large crabs, except in the few places where crabs are naturally rare. Eliminating predation on seed clams by shore crabs is likely impossible using netting, since mesh size would have to be <1 cm to keep out the small crabs that are capable of consuming 2 mm clam seed. A viable alternative may be raising seed in hatcheries until they at least 10-15 mm, achieving a size refuge from shore crabs. This relatively large 'safe' size may be adequate for various thick-shelled clams (Walton, MacKinnon, Rodriguez, Proctor, Ruiz, 2002) but may need to be larger (>20 mm) when dense blue crabs are present (Peterson, Summerson, Huber, 1995). 'Safe' seed size for softshell clams is hard to calculate because they are so readily crushed, or even peeled through their siphonal gape (Boulding, 1984). For shore crabs, farming on beaches lacking the cobble or debris that provide crab habitat may help reduce juvenile clam mortality.

Predation by crabs thus is a multifaceted problem for the valuable shellfish aquaculture industry, especially when the crabs are diverse in their sizes and behaviors, as is true on the tide flats in Washington. The continued invasion of the green crab *Carcinus maenas* into the inside waters of Washington (Grason, McDonald, Adams, Litle, Apple, Pleus, 2018; Yamada, Thomson, Gillespie, Norgard, 2017) will further exacerbate the challenge of clam losses to crab predators.

5. Acknowledgements

We thank the University of Washington and the Friday Harbor Labs for the use of facilities. Dara Yiu, Julia Kobelt, Stephanie Crofts, and Adam P. Summers provided assistance in the lab and field. Access to field sites was generously provided by Taylor Shellfish, Hama Hama Oyster

Company, Heckes Clam and Oyster Company, and the Samish Nation. Nyle and Brittany Taylor provided juvenile clams and invaluable advice. M.Y. and A.M. thank 2017 FHL students for assistance, and the UW Mary Gates Research Scholarship for financial support. This publication was funded by a grant from Washington Sea Grant, University of Washington, pursuant to National Oceanic and Atmospheric Administration Award No. NA14OAR4170078. The views expressed herein are those of the authors and do not necessarily reflect the views of NOAA or any of its sub-agencies.

6. References

- Altieri, A.H., 2008. Dead zones enhance key fisheries species by providing predation refuge. *Ecol.* 89, 2808-2818.
- Arnold, W., 1984. The effects of prey size, predator size, and sediment composition on the rate of predation of the blue crab, *Callinectes sapidus* Rathbun, on the hard clam, *Mercenaria mercenaria* (Linne). *J. Exp. Mar. Biol. Ecol.* 80, 207-219.
- Beal, B.F., 2006. Relative importance of predation and intraspecific competition in regulating growth and survival of juveniles of the soft-shell clam, *Mya arenaria* L., at several spatial scales. *J. Exp. Mar. Biol. Ecol.* 336, 1-17.
- Beal, B.F., Vencile, K., 2001. Short-term effects of commercial clam (*Mya arenaria* L.) and worm (*Glycera dibranchiata* Ehlers) harvesting on survival and growth of juveniles of the soft-shell clam. *J. Shellfish Res.* 20, 1145-1157.
- Becker, P., Barringer, C., Marelli, D., 2008. Thirty years of sea ranching Manila clams (*Venerupis philippinarum*): successful techniques and lessons learned. *Reviews in Fisheries Science.* 16, 44-50.
- Bendell, L.I., 2015. Favored use of anti-predator netting (APN) applied for the farming of clams leads to little benefits to industry while increasing nearshore impacts and plastics pollution. *Mar. Pollut. Bull.* 91, 22-28.
- Beukema, J.J., Dekker, R., 2005. Decline of recruitment success in cockles and other bivalves in the Wadden Sea: possible role of climate change, predation on postlarvae and fisheries. *Mar. Ecol. Prog. Ser.* 287, 149-167.
- Beukema, J.J., Dekker, R., 2014. Variability in predator abundance links winter temperatures and bivalve recruitment: correlative evidence from long-term data in a tidal flat. *Mar. Ecol. Prog. Ser.* 513, 1-15.

281 Beukema, J.J., Dekker, R., Philippart, C.J.M., 2010. Long-term variability in bivalve recruitment,
 282 mortality, and growth and their contribution to fluctuations in food stocks of shellfish-
 283 eating birds. *Mar. Ecol. Prog. Ser.* 414, 117-130.
 284 Boulding, E.G., 1984. Crab-resistant features of shells of burrowing bivalves: decreasing
 285 vulnerability by increasing handling time. *J. Exp. Mar. Biol. Ecol.* 76, 201-223.
 286 Boulding, E.G., LaBarbera, M., 1986. Fatigue damage: repeated loading enables crabs to open
 287 larger bivalves. *Biol. Bull.* 171, 538-547.
 288 Dethier, M.N., Schoch, G.C., 2005. The consequences of scale: assessing the distribution of
 289 benthic populations in a complex estuarine fjord. *Est. Coastal Shelf Sci.* 62, 253-270.
 290 Dethier, M.N., Ruesink, J.L., Berry, H., Sprenger, A.G., 2012. Decoupling of recruitment from
 291 adult clam assemblages along an estuarine shoreline. *J. Exp. Mar. Biol. Ecol.* 422, 48-54.
 292 Dethier, M.N., Kobelt, J.N., Yiu, D., Wentzel, L., Ruesink, J.L., 2019. Context-dependence of
 293 abiotic and biotic factors influencing performance of juvenile clams. *Est. Coastal Shelf*
 294 *Sci.* 219, 201-209.
 295 Glaspie, C., Longmire, K., Seitz, R.D., 2017. Acidification alters predator-prey interactions of blue
 296 crab *Callinectes sapidus* and soft-shell clam *Mya arenaria*. *J. Exp. Mar. Biol. Ecol.* 489,
 297 58-65.
 298 Glaspie, C., Seitz, R.D., Ogburn, M., Dungan, C., Hines, A., 2018. Impacts of habitat, predators,
 299 recruitment, and disease on soft-shell clams *Mya arenaria* and stout razor clams *Tagelus*
 300 *plebeius* in Chesapeake Bay. *Mar. Ecol. Prog. Ser.* 603, 117-133.
 301 Grason, E., McDonald, P.S., Adams, J., Litle, K., Apple, J.K., Pleus, A., 2018. Citizen science
 302 program detects range expansion of the globally invasive European green crab in
 303 Washington State (USA). *Management of Biological Invasions.* 9, 39-47.
 304 Green, M.A., Waldbusser, G.G., Reilly, S.L., Emerson, K., O'Donnell, S., 2009. Death by
 305 dissolution: sediment saturation state as a mortality factor for juvenile bivalves. *Limnol.*
 306 *Oceanogr.* 54, 1037-1047.
 307 Grosholz, E.D., Ruiz, G.M., Dean, C.A., Shirley, K.A., Maron, J.L., Connors, P.G., 2000. The
 308 impacts of a nonindigenous marine predator in a California bay. *Ecol.* 81, 1206-1224.
 309 Harger, J.R., 1972. Competitive coexistence among intertidal invertebrates. *Amer. Sci.* 60, 600-
 310 607.
 311 Higgins, C., Stephenson, K., Brown, B., 2010. Nutrient bioassimilation capacity of aquacultured
 312 oysters: quantification of an ecosystem service. *Environmental Quality.* 40, 271-277.
 313 Hunt, H.L., Scheibling, R.E., 1997. Role of early post-settlement mortality in recruitment of
 314 benthic marine invertebrates. *Mar. Ecol. Prog. Ser.* 155, 269-301.
 315 Hunt, H.L., Mullineaux, L.S., 2002. The roles of predation and postlarval transport in recruitment
 316 of the soft shell clam (*Mya arenaria*). *Limnol. Oceanogr.* 47, 151-164.
 317 Knudsen, J.W., 1964. Observations of the reproductive cycles and ecology of the common
 318 Brachyura and crablike Anomura of Puget Sound, Washington. *Pac. Sci.* 18, 3-33.
 319 Kozloff, E.N., 1993. *Seashore Life of the Northern Pacific Coast*. University of Washington Press,
 320 Seattle.
 321 Liu, D., Wang, F., Lu, Y., Hu, N., Su, X., Liu, J., Zhu, B., 2019. Behavioral mechanisms of swimming
 322 crab (*Portunus trituberculatus*) preying on Manila clams (*Ruditapes philippinarum*):
 323 effects of substrate and competition. *Mar. Biol.* 166.

- Low, C.J., 1970. Factors affecting the distribution and abundance of two species of beach crab, *Hemigrapsus oregonensis* and *Hemigrapsus nudus*, Zoology. University of British Columbia, Vancouver, B.C.
- Munroe, D., McKinley, R.S., 2007. Commercial Manila clam (*Tapes philippinarum*) culture in British Columbia, Canada: the effects of predator netting on intertidal sediment characteristics. Est. Coastal Shelf Sci. 72, 319-328.
- Munroe, D., Kraeuter, J., Beal, B.F., Chew, K.K., Luckenbach, M., Peterson, C.P., 2015. Clam predator protection is effective and necessary for food production. Mar. Pollut. Bull. 100, 47-52.
- Myers, R., Baum, J., Shepherd, T., Powers, S.P., Peterson, C., 2007. Cascading effects of the loss of apex predatory sharks from a coastal ocean. Science. 315, 1846-1850.
- Olafsson, E.B., 1989. Contrasting influences of suspension-feeding and deposit-feeding populations of *Macoma balthica* on infaunal recruitment. Mar. Ecol. Prog. Ser. 55, 171-179.
- Peterson, C., Summerson, H., Huber, J., 1995. Replenishment of hard clam stocks using hatchery seed: combined importance of bottom type, seed size, planting season, and density. J. Shellfish Res. 14, 293-300.
- Peterson, C., Grabowski, J., Powers, S.P., 2003. Estimated enhancement of fish production resulting from restoring oyster reef habitat: quantitative valuation. Mar. Ecol. Prog. Ser. 264, 249-264.
- Quayle, D.B., 1988. Pacific oyster culture in British Columbia. Canadian Bulletin of Fisheries and Aquatic Sciences. 218, 1-241.
- Ruesink, J.L., Freshley, N., Herrold, S., Trimble, A.C., Patten, K., 2014. Influence of substratum on non-native clam recruitment in Willapa Bay, Washington, USA. J. Exp. Mar. Biol. Ecol. 459, 23-30.
- Saurel, C., Ferreira, J., Cheney, D.P., Suhrbier, A.D., Dewey, B., Davis, J., Cordell, J.R., 2014. Ecosystem goods and services from Manila clam culture in Puget Sound: a modelling analysis. Aquaculture Environment Interactions. 5, 255-270.
- Smith, M., Langdon, C., 1998. Manila clam aquaculture on shrimp-infested mudflats. J. Shellfish Res. 17, 223-229.
- Tezuka, N., Kamimura, S., Hamaguchi, M., Saito, H., Iwano, H., Egashira, J., Fukuda, Y., Tawaratsumida, T., Nagamoto, A., Nakagawa, K., 2012. Settlement, mortality and growth of the asari clam (*Ruditapes philippinarum*) for a collapsed population on a tidal flat in Nakatsu, Japan. J. Exp. Mar. Biol. Ecol. 439, 108-112.
- Walton, W.C., MacKinnon, C., Rodriguez, L.F., Proctor, C., Ruiz, G.M., 2002. Effect of an invasive crab upon a marine fishery: green crab, *Carcinus maenas*, predation upon a venerid clam, *Katelysia scalarina*, in Tasmania (Australia). J. Exp. Mar. Biol. Ecol. 272, 171-189.
- Whitton, T., Jenkins, S., Richardson, C., Hiddink, J., 2012. Aggregated prey and predation rates: juvenile shore crabs (*Carcinus maenas*) foraging on post-larval cockles (*Cerastoderma edule*). J. Exp. Mar. Biol. Ecol. 432, 29-36.
- Williams, J.G., 1980. Growth and survival in newly settled spat of the Manila clam, *Tapes japonica*. Fish. Bull. 77, 891-900.
- Yamada, S.B., Boulding, E.G., 1996. The role of highly mobile crab predators in the intertidal zonation of their gastropod prey. J. Exp. Mar. Biol. Ecol. 204, 59-83.

368 Yamada, S.B., Boulding, E.G., 1998. Claw morphology, prey size selection and foraging efficiency
369 in generalist and specialist shell-breaking crabs. J. Exp. Mar. Biol. Ecol. 220, 191-211.
370 Yamada, S.B., Metcalf, H., Baldwin, B.C., 1993. Predation by the crab, *Cancer oregonensis* Dana,
371 inside oyster trays. J. Shellfish Res. 12, 89-92.
372 Yamada, S.B., Thomson, R.E., Gillespie, G.E., Norgard, T.C., 2017. Lifting barriers to range
373 expansion: the European green crab *Carcinus maenas* (Linnaeus, 1758) enters the Salish
374 Sea. J. Shellfish Res. 36, 1-8.
375

FIGURE CAPTIONS

Figure 1. Relative sizes of crabs (*Hemigrapsus* spp.) and clams in trials where crabs were able to consume the clams offered. N = 33 trials with Manila clams, N = 20 trials with *Mya*. R² values of linear regression for each clam species: *Mya* 0.20, Manila 0.46.

Figure 2. Predation rates by crabs on Manila clams of varying sizes. Crab Dactyl sizes: Small ≤ 4 mm; Medium = 4.1 to 6.0; Large = 6.1 to 8.0 mm. Bars are mean and one s.e. of N trials; replicate N's per bar (left to right) = 130, 0, 0, 41, 12, 8, 19, 14, 18. No trials were run (ND = no data) attempting to feed medium or large clam categories to small crabs.

Figure 1.

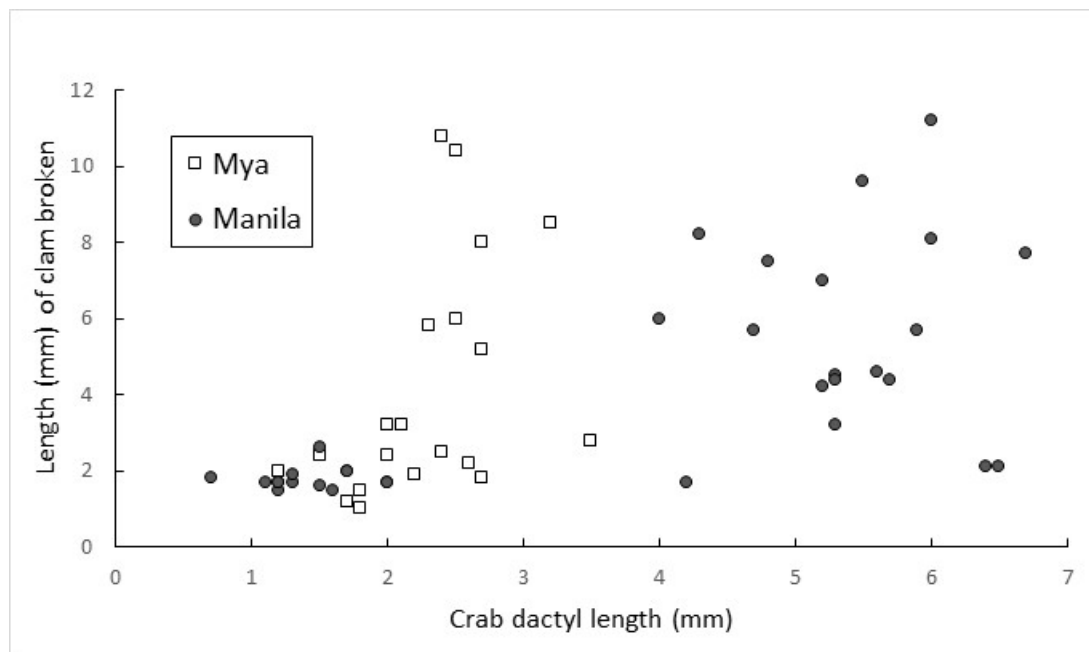
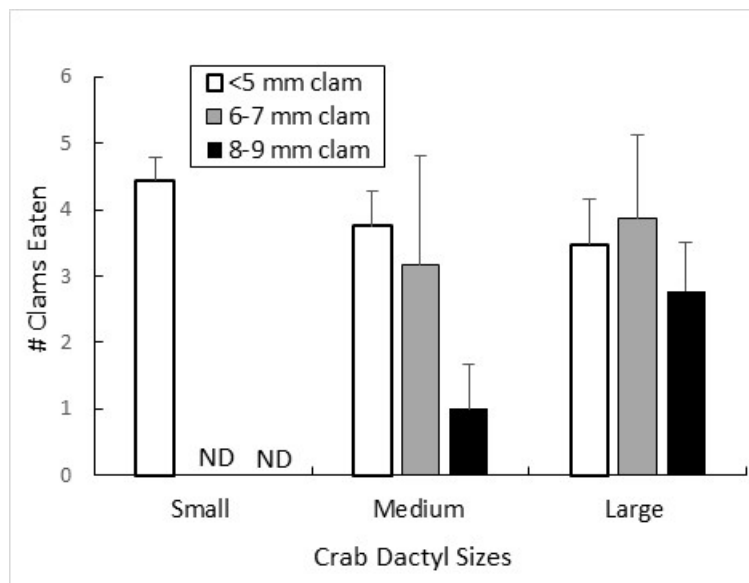
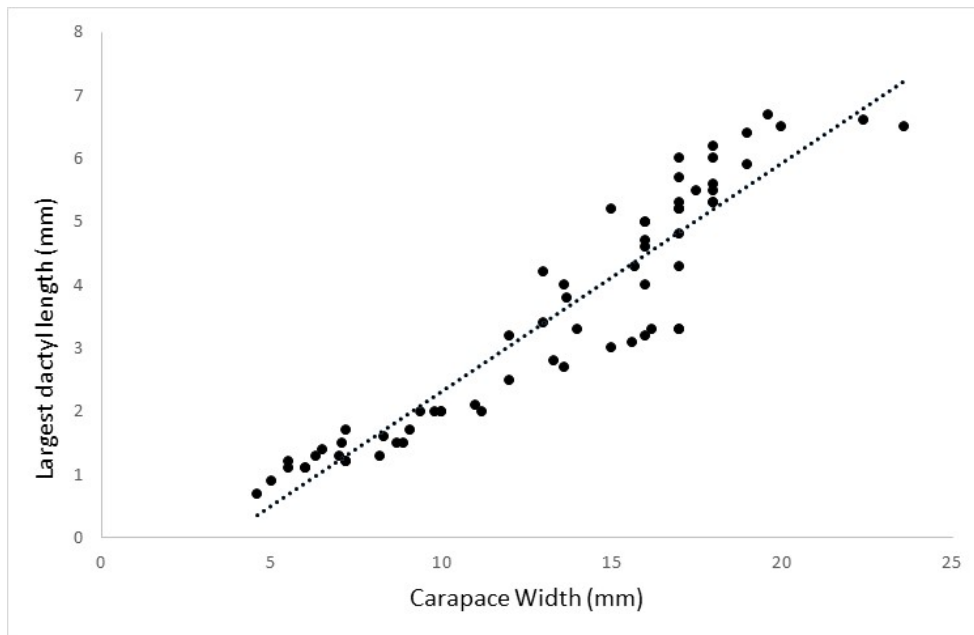
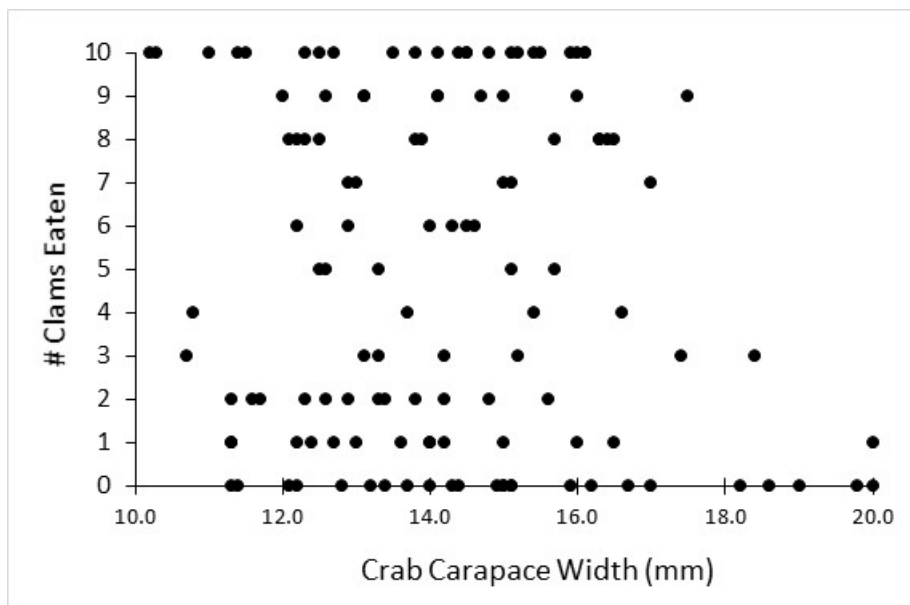


Figure 2.

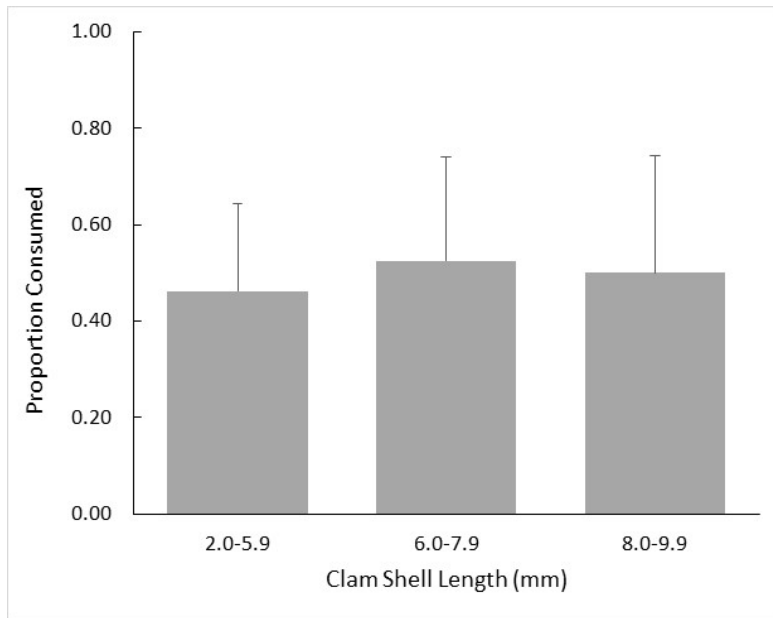




Supplemental Figure 1. Correlation between carapace size and dactyl length for *Hemigrapsus oregonensis*. N = 66 crabs, $r^2 = 0.88$



Supplemental Figure 2. Numbers of small (2 mm) clams eaten out of 10 offered to different sizes of *Hemigrapsus* spp. over 24 hours. N = 126 trials, each point is a separate individual crab.



Supplemental Figure 3. Mean (one s.d.) proportion of clams consumed by shore crabs when offered a choice of 5 clams of each size class. N=16 trials (separate crabs).